

In Vitro Synergistic Effect of Xylitol with *Salvadora persica* L Extracts and Cephalixin on *Streptococcus mutans* Strains

Salah G. Ali*

Botany and Microbiology Department, faculty of Science, Al-Azhar University, 71524 Assiut, EGYPT

Received: 4 Feb. 2017, Revised: 6 April. 2017, Accepted: 10 April. 2017.

Published online: 1 May 2017.

Abstract: Sixty six *Streptococcus mutans* isolates have been tested with cephalixin and *Salvadora persica* L extracts with or without addition of glucose, or xylitol to determine minimal inhibitory concentration (MICs), susceptibility of *S. mutans* isolates and time-killing curve. Result: All the strains were sensitive to cephalixin (12.8. ug/ml), *S. persica* aqueous extract (16.0 mg/ml), *S. persica* methanol extract (4.0 mg/ml), *S. persica* ethanol extract (12.0 mg/ml) and *S. persica* hexane extract (8.0 mg/ml). The addition of xylitol increased susceptibility of *S. mutans* isolates to the MIC values of cephalixin and *S. persica* extracts, whereas the addition of glucose decreased susceptibility of *S. mutans* to the MIC values of cephalixin and *S. persica* extracts. Time-killing curve of cephalixin combined with xylitol exhibit zero CFU/ml after 16 hrs. and xylitol combined with *S. persica* hexane extract exhibit zero CFU/ml after 18 hrs with highly significant values. Conclusion: The mix of xylitol with *S. persica* extracts or cephalixin may lead to inhibit microbial causative agents of dental decay, while addition of glucose has opposite effect.

Keywords: *Streptococcus mutans*, *Salvadora persica*, cephalixin.

1 Introduction

Dental decay is due to the dissolution of tooth mineral by acids derived from bacterial fermentation of sucrose and other dietary carbohydrates. *Streptococcus mutans* is naturally present in the human oral microbiota, along with at least 25 other species of oral streptococci and consider the main cause of dental decay [18,5]. *Streptococcus mutans* characterized by it's a Gram-positive, non-motile, non-spore forming, facultative anaerobic cocci bacterium commonly found in the human oral cavity [7]

Good oral and dental hygiene can be achieved by avoiding sugary sweets and regular brushing to avoid tooth decay.

Salvadora persica L, has important role in the oral hygiene [31], because it has many antimicrobial agents such as flavonoid [14], glycosides [15], fluoride [10], sulfur-containing organic substances [21], and several anionic components [14]

Sucrose is the only sugar that bacteria can use to form this sticky polysaccharide [25], while glucose sugar fermented by *S. mutans* strains to produce lactic acid [19]. On the other hand, fermentable sucrose sugar was substituted by non-fermentable xylitol sugar for growth inhibition of *Streptococcus mutans* [6]. Xylitol, a naturally in fruit, vegetables, and berries and is artificially manufactured from xylan-rich plant materials [22, 3].

This research aimed to determine the effect of antimicrobial activity of *Salvadora persica* L extracts and cephalixin with or without addition of xylitol against *Streptococcus mutans* isolates.

2 Materials and Methods

2.1 *Streptococcus* Isolation And Identification Method

Streptococcus sp were isolated from buccal surfaces of the caries teeth of patients of the general dental clinic (Faculty of dentistry, Al-Azhar University, Assiut branch, EGYPT) were grown on MS (mitis-salivarius agar) plates and incubated anaerobically, using anaerobic candle jar, for 48 hrs. at 37C. Count of more than 250 colonies (10⁴cells/ml) was considered as positive samples [12,11]. Colonies grown on MS-agar medium was transferred and purified on the blood agar plates and incubated anaerobically for two days. The identification of *S. mutans* according to Bergeys Manual of Determinative Bacteriology 9th., 1994. Confirmation of the identification by using commercial kit ApI20 strep and Dextran Production Test [13].

*Corresponding author e-mail: salahalibadr@yahoo.com

2.2 Cephalixin

Cephalixin Sigma-Aldrich -C4895 was used in this study.

2.3 Xylitol And Glucose

Sugar used in this study glucose and xylitol where purchased from Sigma-Aldrich Co.

2.4 Plant Samples

S. persica were purchased from local market at Assiut city, EGYPT. *S. persica* imported to Egypt from Saudi Arabia. (AlHaramin Company)

2.5 Preparation Of *S Persica* Extracts

The air-dried plant materials were ground into fine powder in grinder and extracted with distilled water, methanol 80%, ethanol 95% and hexane. A 100g sample of ground plant was soaked in 500 ml solvent. The extract was filtered through a Buchner funnel with Whatman filter paper, after filtration of total extracts; the extracts were evaporated under reducing pressure to dryness at 45 °C on a rota-evaporator (Büchi R114), all extracts were soluble in dimethylformaamid (DMF) with exception of the aqueous extracts dissolved in distilled water.

2.6 Determination Of Minimal Inhibitory Concentration (MIC) [4]

An agar dilution method used to determine the minimal inhibitory concentration (MIC) for cephalixin and all *S. persica* different extracts.. Serial concentrations of all tested extracts were achieved (% v/v) in plates containing BHI agar medium. Petri plates of BHI agar containing various concentrations of cephalixin or *S. persica* extracts were inoculated with 24 h culture of bacterial strain by spreading using glass rod in triplicates Each antibacterial test also included plates containing the culture medium plus solvent to obtain a control of the solvent antimicrobial effect. After inoculation procedures, using triplicates, test plates and control plate were then incubated at 37°C anaerobically using anaerobic candle jar in presence of 5% CO₂. Plates were evaluated for the presence or absence of visible growth of each strain after 24, 48 and 72 hrs of incubation. The absence of colonies on all plates tested was considered as an inhibitory effect. The lowest concentration of cephalixin or extracts required to inhibit the growth of the tested microorganism completely was designated as the MIC.

2.7 Synergistic Effect Of Xylitol With Cephalixin And *S persica* L Extracts [4]

The cells were cultured in 5ml Brain Heart Infusion (BHI) overnight at 37 °C. The cells were transferred to fresh BHI. Sets of plates contain 1% concentration of glucose with *S persica* L extracts, set of plates contain xylitol with *S persica* L extracts, set of plates contain glucose with cephalixin, and set of plates contain xylitol with cephalixin. After inoculation procedures, using triplicates, test plates and control plate were then incubated at 37°C anaerobically using anaerobic candle jar in presence of 5% CO₂. *In vitro* synergism assays of xylitol in combination with cephalixin and/ or with different *S. persica* L extracts were carried out after evaluating the MIC of all test different extracts on BHI agar medium by agar dilution method.

2.8 Assessment Of Killing Curve Of Xylitol With Cephalixin And Hexane *S Persica* L Extract [4]

Selected strains of *Streptococcus mutans* isolates were grown overnight in BHI broth medium. 0.2 ml of inoculum was added to 20 ml BHI flasks containing antibacterial agent 4 MIC for cephalixin plus 1% xylitol, (1MIC) for *S. persica* L hexan extract, plus 1% xylitol in combination. Flasks were then incubated at 37°C using anaerobic candle jar in presence of 5% CO₂. After 2 hrs. the flasks were strongly agitated and a 0.1ml sample was diluted and plated; and the flasks were immediately returned to incubation.

2.9 Statistical Analysis

All experiments were carried out with three replicates. Statistical tests Analysis of variance (ANOVA) was performed.

3 Results

3.1 Determination Of Antibacterial Activities Of Cephalixin Against *S. Mutans* Isolates

According to the present results, the minimal inhibitor concentration of cephalixin on *Streptococcus mutans* ranged between 0.4 and 12.8 µg/ml where results exhibited 6 isolates sensitive for 0.4 µg/ml, whereas MIC for 10 isolates was 0.8 µg/ml, 16 isolates MIC was 1.6 µg/ml, 34 isolates MIC was 3.2 µg/ml, 47 isolates MIC was 6.4 µg/ml, and 66 isolates was sensitive to 12.8 µg/ml of cephalixin compared with cephalixin plus glucose where number of susceptible isolates decreased. Highest increasing ratio of susceptibility by addition of xylitol was at 0.8 µg/ml of cephalixin by 62.1% while addition of 1% glucose decreased susceptibility of isolates by 21.2% at 6.4

µg/ml of cephalixin (Table 1., Figure 1)

3.2 Determination of antibacterial activities of extracts of *S. presica L*

According to the present results, the minimal inhibitory concentration of *Salvadora presica L* extracts on *Streptococcus mutans* isolates ranged between 6 and 16 mg /ml of aqueous extract, 2 and 4mg /ml of methanol extract , 6 and 12 mg /ml of ethyl alcohol extract , while hexane extract exhibited 4 and 8 mg /ml. Number of susceptible isolates varied according concentration of extracts and solvent types, where *S. presica L* aqueous extract exhibited 15 isolates sensitive for 4 mg/ml, whereas MIC for 20 isolates were 8 mg/ml, and all 66 isolates were sensitive to 16 mg/ml. *S. presica L* methanol extract exhibited five isolates sensitive for 2 mg/ml,

whereas 33 isolates were sensitive to 3 mg/ml, and all 66 isolates were sensitive to 4 mg/ml. *S. presica L* ethanol extract exhibited twelve isolates were sensitive to 6 mg/ml, whereas 16 isolates were sensitive to 8 mg/ml, and all 66 isolates were sensitive to 12 mg/ml. *S. presica L* hexane extract exhibited eight isolates were sensitive to 4 mg/ml, whereas 17 isolates were sensitive to 6 mg/ml, and all 66 isolates were sensitive to 8 mg/ml.

By addition of 1% glucose into *Salvadora presica L* extracts in mixture exhibited decreasing *Streptococcus mutans* susceptibility numbers at all concentration, whereas by addition of 1% xylitol exhibited increasing of susceptibility numbers of *Streptococcus mutans*, where the number of susceptibility increased by 54.5% at 6mg/ml of SHE, followed by 25.8% at 8mg/ml SEE, 19.7 % at 8 mg/ml SEE and 16.7% at 3 mg/ml SME (Table 2, figures 2, 3, 4 & 5)

Table 1. Antibacterial activities of cephalixin, cephalixin with glucose & cephalixin with xylitol against 66 *Streptococcus mutans* isolates

| Antibiotic | Susceptibility | Concentration (µ g /ml) | | | | | |
|------------|-----------------------------|-------------------------|------|------|------|------|------|
| | | 0.4 | 0.8 | 1.6 | 3.2 | 6.4 | 12.8 |
| Ce | No. of susceptible strain | 6 | 10 | 16 | 34 | 47 | 66 |
| | % No. Of susceptible strain | 9.0 | 15.2 | 24.2 | 51.5 | 71.2 | 100 |
| Ce -g 1% | No. of susceptible strain | 2 | 6 | 8 | 27 | 33 | 59 |
| | % No. Of susceptible strain | 3 | 9.1 | 12.1 | 40.9 | 50 | 89.4 |
| Ce -X 1% | No. of susceptible strain | 40 | 51 | 55 | 60 | 62 | 66 |
| | % No. Of susceptible strain | 60.6 | 77.3 | 83.3 | 90.9 | 93.9 | 100 |

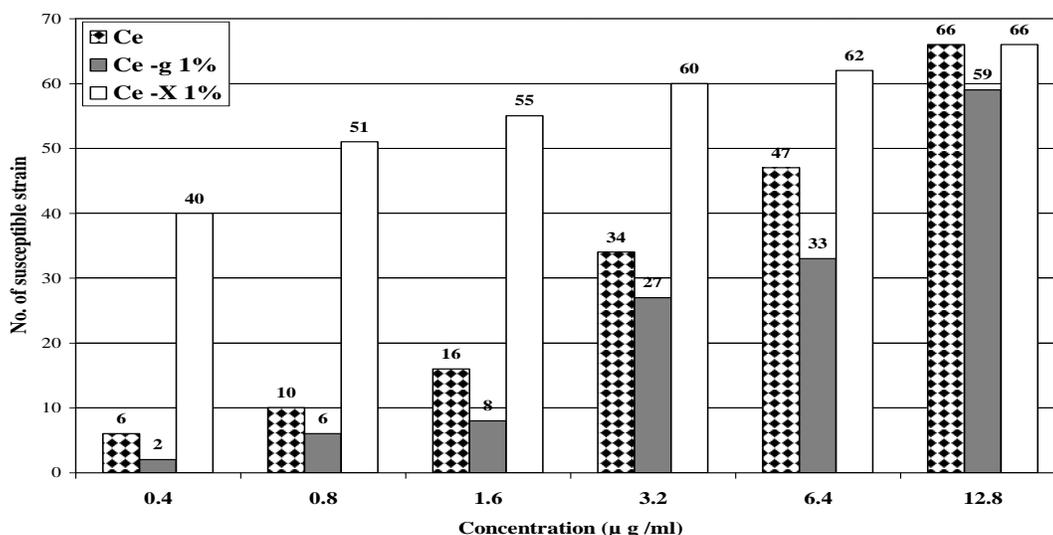


Fig1. Number of *Streptococcus mutans* susceptible stains to cephalixin Ce, Ce-g & Ce-x

Table 2. Antibacterial activities of extracts of *S. presica*, *S. presica* extracts with glucose and extracts of *S. presica* with xylitol against 66 *Streptococcus mutans* strains

| <i>S. presica</i> aqueous extract (SEE) | | | | |
|--|---------------------------|------|------|------|
| SAE | Concentration (mg/ml) | 4 | 8 | 16 |
| | No. of susceptible strain | 15 | 20 | 66 |
| | (%) susceptible strain | 22.7 | 30.3 | 100 |
| SAE-G 1% | No. of susceptible strain | 12 | 14 | 55 |
| | (%) susceptible strain | 18.2 | 21.2 | 83.4 |
| SAE-X 1% | No. of susceptible strain | 27 | 33 | 66 |
| | (%) susceptible strain | 40.9 | 50 | 100 |
| <i>S. presica</i> methanol extract (SME) | | | | |
| SME | Concentration (mg/ml) | 2 | 3 | 4 |
| | No. of susceptible strain | 5 | 33 | 66 |
| | (%) susceptible strain | 7.8 | 50 | 100 |
| SME-G 1% | No. of susceptible strain | 2 | 12 | 50 |
| | (%) susceptible strain | 3 | 18.2 | 75.7 |
| SME-X 1% | No. of susceptible strain | 9 | 44 | 66 |
| | (%) susceptible strain | 13.6 | 66.7 | 100 |
| <i>S. presica</i> Ethanol extract (SEE) | | | | |
| SEE | Concentration (mg/ml) | 6 | 8 | 12 |
| | No. of susceptible strain | 12 | 16 | 66 |
| | (%) susceptible strain | 18.2 | 24.2 | 100 |
| SEE-G 1% | No. of susceptible strain | 8 | 14 | 56 |
| | (%) susceptible strain | 12.1 | 21.2 | 84.8 |
| SEE-X 1% | No. of susceptible strain | 25 | 33 | 66 |
| | (%) susceptible strain | 37.9 | 50 | 100 |
| <i>S. presica</i> Hexane extract (SHE) | | | | |
| SHE | Concentration (mg/ml) | 4 | 6 | 8 |
| | No. of susceptible strain | 8 | 17 | 66 |
| | (%) susceptible strain | 12.1 | 27.3 | 100 |
| SHE-g 1% | No. of susceptible strain | 5 | 12 | 55 |
| | (%) susceptible strain | 7.6 | 18.2 | 83.3 |
| SHE-X 1% | No. of susceptible strain | 29 | 53 | 66 |
| | (%) susceptible strain | 43.9 | 80.3 | 100 |

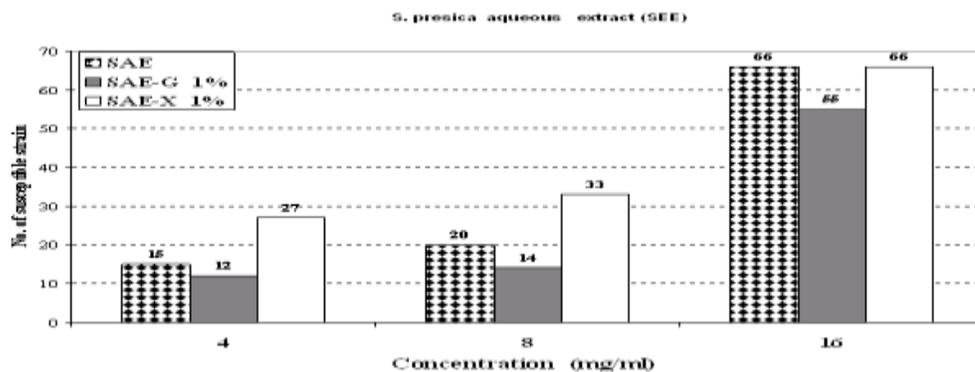


Fig 2. Number of *Streptococcus mutans* susceptible stains to SAE, SAE-g & SAE-x

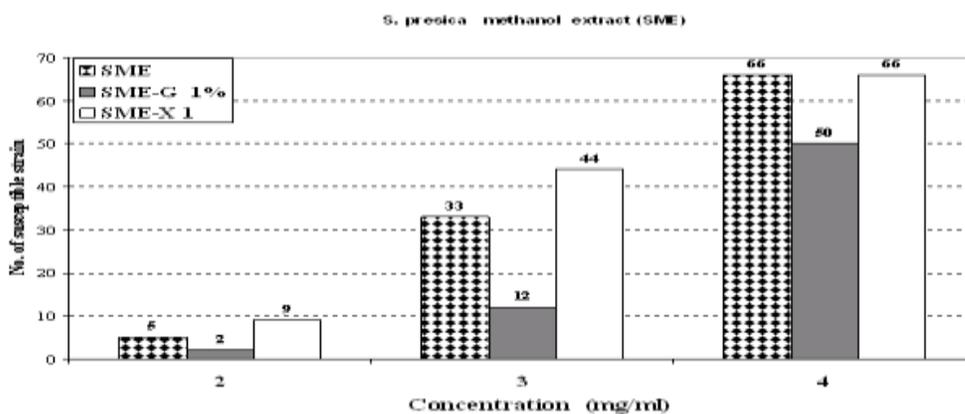


Fig 3. Number of *Streptococcus mutans* susceptible stains to SME, SME-g & SME-x

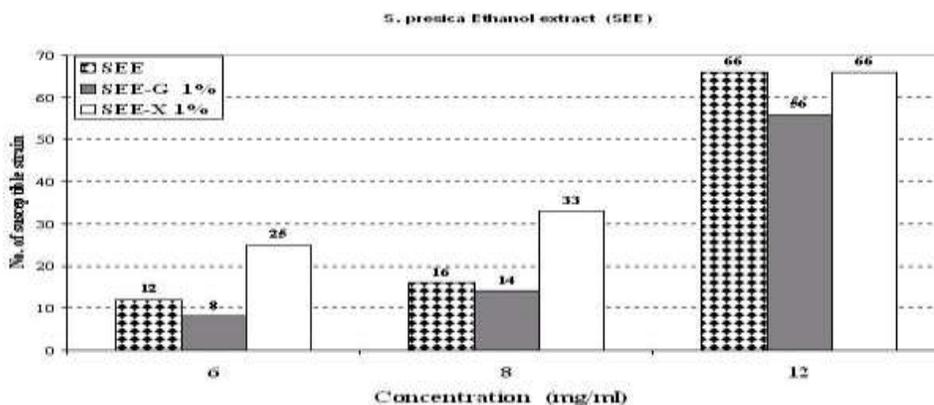


Fig 4. Number of *Streptococcus mutans* susceptible stains to SEE, SEE-g & SEE-x

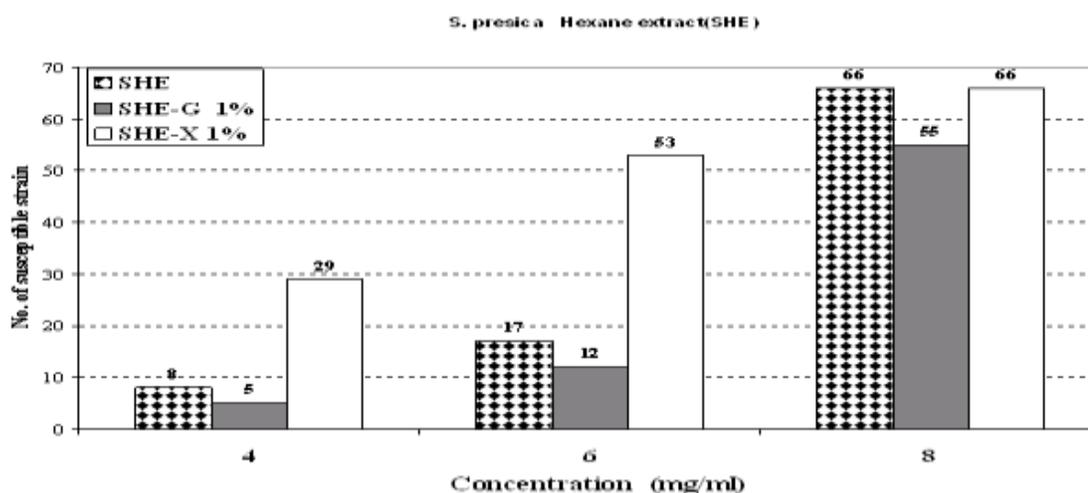


Fig 5. Number of *Streptococcus mutans* susceptible stains to SHE, SHE-g & SHE-x

3.3. Time-Killing Curve Of Cephalexin And *S. presica* Hexane Extract (SHE) With Or Without Addition Of 1% Glucose Or 1% Xylitol Against *S. Mutans* Strains.

This research study has only focused on time-killing curve of cephalexin and *S. presica* hexane extract (SHE)

with or without addition of 1% glucose or 1% xylitol challenged against selected *S. mutans* strains.

Cephalexin exhibit zero CFU/ml at 20 hrs of incubation time while cephalexin with xylitol exhibited zero CFU/ml after 16 hrs of incubation period whereas cephalexin with 1% glucose, *S. mutans* exhibited resistance to cephalexin (Table 3. Fig 6)

Table 3. Time of killing rate (log 10 CFU/ 0.1 ml) of cephalexin (c) against *Streptococcus mutans* isolates compared with cephalexin with 1% glucose (cg) & cephalexin with 1% xylitol (cx) compared with growth curve (G)

| Time | G | C | Cg | Cx | F-test | L.S.D. |
|------|------|------|------|------|--------|--------|
| 0 | 6.64 | 6.64 | 6.64 | 6.64 | n.s | ---- |
| 4 | 7.75 | 5.54 | 6.81 | 4.64 | ** | 0.065 |
| 6 | 7.90 | 4.52 | 6.83 | 3.99 | ** | 0.058 |
| 8 | 8.29 | 3.82 | 6.85 | 2.54 | ** | 0.095 |
| 10 | 8.44 | 3.74 | 6.85 | 1.99 | ** | 0.121 |
| 12 | 8.99 | 2.54 | 6.81 | 1.74 | ** | 0.115 |
| 16 | 8.98 | 2.39 | 6.71 | 0 | ** | 0.175 |
| 18 | 8.94 | 1.54 | 6.65 | 0 | ** | 0.191 |
| 20 | 7.88 | 0 | 6.58 | 0 | ** | 0.123 |
| 22 | 6.81 | 0 | 6.51 | 0 | ** | 0.107 |
| 24 | 6.79 | 0 | 5.88 | 0 | ** | 0.121 |

*=Significant **=highly significant ns= no significant

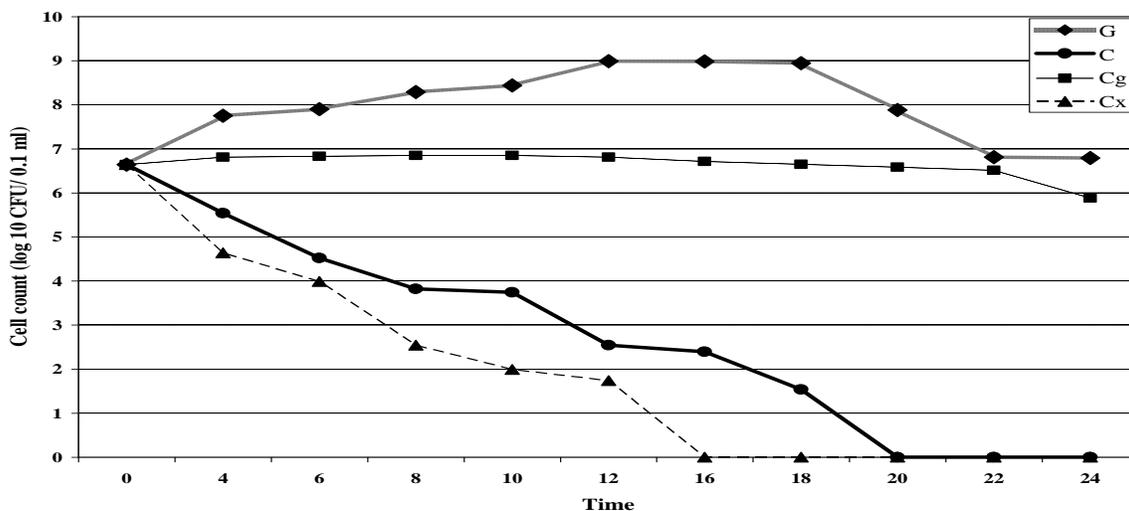


Fig 6. Time of killing rate (log 10 CFU/ 0.1 ml) of cephalixin (c) against *Streptococcus mutans* isolates compared with Cg, Cx & growth curve (G)

The effect of xylitol against *S. mutans* indicated number of *S. mutans* inoculant not increased. i.e. bacteriostatic effect comparing with growth curve without any treatment. *S. presica* hexane extract (SHE) exhibited zero CFU after 22

hrs. of incubation time whereas by addition of 1% xylitol to SHE exhibited zero CFU after 18 hrs. of incubation period with highly significant values., while glucose addition to SHE exhibit resistance effect to activity of the extract (Table 4, Fig. 7)

Table 4. Time of killing rate (log 10 CFU/ 0.1 ml) of *S. presica* hexane extract (SH) against *Streptococcus mutans* isolates compared with SHE with 1% glucose (SH-g), SH with 1% xylitol (SH-x) & xylitol 1% (X)

| Time | X | SH | SHg | SHx | F-test | L.S.D. |
|------|------|------|------|------|--------|--------|
| 0 | 6.64 | 6.64 | 6.64 | 6.64 | ns | ---- |
| 4 | 6.49 | 6.54 | 6.85 | 5.99 | * | 0.045 |
| 6 | 5.43 | 6.52 | 6.85 | 4.52 | ** | 0.048 |
| 8 | 5.20 | 6.46 | 6.74 | 3.82 | ** | 0.105 |
| 10 | 5.07 | 5.99 | 6.85 | 2.39 | ** | 0.117 |
| 12 | 5.07 | 5.94 | 6.81 | 2.20 | ** | 0.174 |
| 16 | 5.06 | 4.46 | 6.71 | 1.65 | ** | 0.165 |
| 18 | 5.07 | 3.94 | 6.65 | 0 | ** | 0.095 |
| 20 | 5.06 | 1.98 | 6.58 | 0 | ** | 0.185 |
| 22 | 5.06 | 0 | 6.51 | 0 | ** | 0.164 |
| 24 | 5.07 | 0 | 5.88 | 0 | ** | 0.054 |

*=Significant ** =highly significant ns= no significant

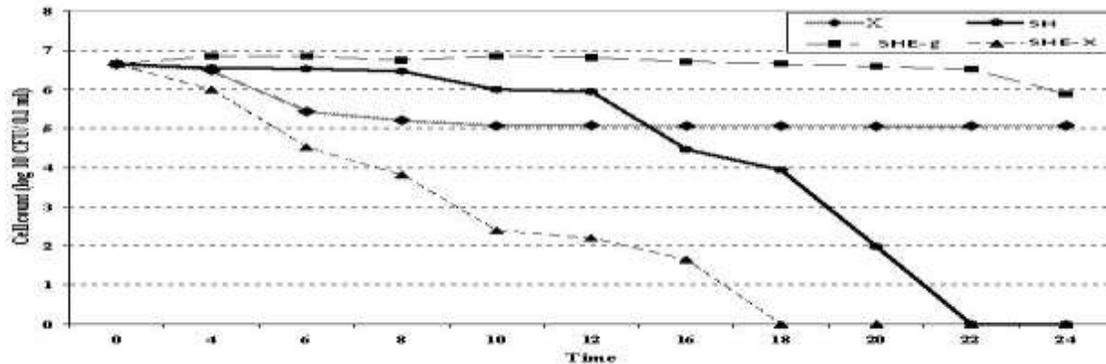


Fig 7. Time of killing rate (log 10 CFU/ 0.1 ml) of SH against *Streptococcus mutans* isolates compared with SHg, SHx and X

4 Discussion

Streptococcus mutans is the primary bacterium involved in plaque formation and the initiation of dental caries and the most cariogenic of all the oral Streptococci responsible for dental caries [18]. Many antimicrobial agents have been used to control effect of *S mutans* involve antibiotics and medicinal plants.

Salvadora persica is considering one of very important traditional tool for oral hygiene. On the other hand xylitol has been used to reduce growth of *S mutans* works as bacteriostatic.

This research paper mainly aimed to test presence of synergistic effect between xylitol and *S presica*, furthermore xylitol with cephalixin antibiotic, which compared with addition effect of glucose only. The beginning of this research aimed to clarify differences in the results of the previous researches regarding antimicrobial activity of *S. presica* L plant against *S. mutans*. The result of this study showed that all *S. presica* extraction solvents (water, methanol, ethanol and hexane) exhibit antimicrobial activity against *S. mutans* with relatively differences according to isolated strains, solvent and concentration of the extract.

Other research results reported that there is no effect of any *Salvadora persica* alcoholic extracts against *S. mutans* [2], otherwise other research study exhibits alcohol is relatively a more efficient solvent than water for the extraction of bioactive compounds of *Salvadora persica* than water [27]. On the other hand, other research study exhibited that the aqueous extract was more potent than alcoholic form in inhibiting microorganism [1]. Those differences in potency of extracted compounds may due to plant from different regions and may due to time of harvesting. Tiwari indicated that botanical source, time of harvesting stage of

development, and method of extraction in addition to the composition, structure, and functional groups of the natural compounds affecting on the antimicrobial activity of natural compounds [29]

The result of this research paper exhibited that addition of glucose to cephalixin and extracts of *S presica* led to reduce susceptibility of *Streptococcus mutans* isolates, that because glucose sugar enhanced isolates ability to grow and because the growth environment is favorable. High environmental glucose concentration led to a decrease in the culture pH and an increase in *S. mutans* growth [32]. Therefore, the MICs can fail to clear higher density infections [17]. Previous study reported that 21% of *Streptococcus mutans* strains were sensitive in the presence of sucrose and resistant in the presence of glucose to cephalosporins [24].

Sugar alcohols such as xylitol were not utilized by oral organisms and may be used as sugar substitutes to reduce dental caries incidence [28]. From the results of this research study, in case of addition of xylitol only within inoculated medium showed that there are reductions of *S. mutans* average numbers.

Streptococcus mutans cannot use alcoholic sugars such as xylitol and sorbitol. Xylitol only absorbed and accumulated in *M. streptococcus* with no energy yield [20]. According the results of this research, addition of xylitol to cephalixin and *S. presica* extracts increased susceptibility of *Streptococcus mutans* isolates at different tested MICs. Wåler and Rølla, reported that xylitol is recommended as a sugar substitute and it has a certain bacteriostatic effect against *S. mutans* and *S. sanguis* [30]. El-Sherbiny indicated that inhibited growth of *S. mutans* depend on xylitol concentration [9]

Other research study exhibited that the cholrhexidin, cationic peptides and xylitol combinations was efficient and superior to single treatments in suppressing *S. mutans* [23], [16]. Other research results indicate that synergistic effect

of xylitol and ursolic acid combination on oral bio films [33], and there are significant reductions in the scores of *S. mutans* were found after the four week use of 20% xylitol mouthrinse [8]. Combination between xylitol with other dental therapies can reduce the incidence of new tooth decay and arrest existing dental caries [22]. According to the results of killing curve of xylitol and *Salvadora persica* hexane extract or with cephalexin combination exhibited clear synergistic effect. Synergy to be present if the number of CFU was $\geq 2 \log_{10}$ lower in the presence of the combination than with the single while antagonism to be present if CFU/0.1ml were $\geq 2 \log_{10}$ higher after incubation with the combination than with the single [26]. According to the obtained results, we recommended that addition of xylitol (bacteriostatic substance) to *S. persica* extracts (bactericidal substances) is considering an important route to overcome tooth decay.

5 Conclusion

In conclusion, both *Salvadora persica* and xylitol have evidence to control *Streptococcus mutans* as synergistic effect, so the addition of xylitol to *Salvadora persica* extract can be a useful method to control and minimize tooth decay by *Streptococcus mutans*. However, it should be one of the choices to increase susceptibility to oral pathogen, to minimize this plant extract concentration and short time for recovery from *Streptococcus mutans* infection. To complete this research study, implementation the final result of this research *in vivo* using xylitol- *Salvadora persica* extracts mixtures necessarily required to make sure that mixtures can be used as a new product of oral hygiene.

Acknowledgement

This research was partially supported by (Al-Ghad International Health Sciences Colleges -AIHSC, Colleges headquartered in Riyadh, Saudi Arabia).

I would also like to show my gratitude to the (Aid Bin Mohamed Al-Aid, professor of Mathematical Science, Qassem University, Al-Ghad International Health Sciences Colleges, Saudi Arabia) for his support and advice during this research.

References

- [1] F. A. Al-Bayati and KD. Sulaiman: In vitro antimicrobial activity of *Salvadora persica* L. extracts against some isolated oral pathogens in Iraq. Turkish J Biol.; 32(1):57–62. 2008.
- [2] K. Almas: Miswak (chewing stick) and its role in oral health, Postgrad. Dent. Middle East 3:214–218, 1993.
- [3] C. Aminoff, E. Vanninen, and T. E. Doty: The occurrence, manufacture and properties of xylitol, In Xylitol, Ed. Counsell, J. N., Applied Science Publishers Ltd, London, pp.1-9. C.F. Microbial production of xylitol, l-xylulose and xylose. Aalto University publication series, DOCTORAL DISSERTATIONS, 2013.
- [4] M. Balouiri, M. Sadiki, and S.K. Ibsouda: Methods for in vitro evaluating antimicrobial activity: A review. Journal of Pharmaceutical Analysis. 71–79, 2016.
- [5] J. A. Banas: Virulence properties of *Streptococcus mutans*. Front. Biosci. 19, 1267–1277. [8], 2004.
- [6] A. Bär : Caries prevention with xylitol. A review of the scientific evidence. World Rev Nutr Diet.;55:183–209, 1988.
- [7] B. U. David, O.O. Linda, and O. E. Charles: Isolation, Characterization and Antibiotic Susceptibility Studies of Clinical Isolates of *Streptococcus mutans* Obtained from Patients Visiting Major Dental Clinics in Nsukka, Nigeria . AJSPSP 2(1): 115, 2011.
- [8] M. ElSalhy, I. Sayed Zahid, and E. Honkala.: Effects of xylitol mouthrinse on *Streptococcus mutans*. J Dent.;40(12):1151-4.doi: 10.1016/j.dent.08.014, 2012.
- [9] M. G. El-Sherbiny: Control of growth *Streptococcus mutans* isolated from saliva and dental caries. Int .J. Curr. Microbiol. App.Sci. 3(10) 1-10, 2014.
- [10] M. Farooqi. And J. Srivastava: The toothbrush tree (*Salvadora persica*) J. Crude Drug Res., 8, pp. 1297-1299, 1968 C.f. Halawany S. H. A review on miswak (*Salvadora persica*) and its effect on various aspects of oral health The Saudi Dental Journal Volume 24, Issue 2, P. 63-69, 2012.
- [11] S. Fingold, and E. Barone: Method of identification of etiological agent of infection disease. In: "Bailey and Scotts diagnostic microbiology". 7th EDT. C. V. Mosby; C. St. Louis. P: 382, 1986.
- [12] J. Friedrich: The genus *Streptococcus mutans* and dental caries. In: "Prokaryotes Hand Book of Habitats, Isolation and Identification of Bacteria ". Mortimer, P.S. (ed.), Barlin, New York, pp. 159801613, 1981.
- [13] O. Guthof, (Cited in Fridrich, J. (1981). The Genus streptococcus and dental disease in: "prokaryotes Hand Book of Habitats, isolation and identification of bacteria ".Mortimer, P. S. (ed.), Berlin, New York, PP. 1598-1613.1970, 1981.
- [14] M. M. Ibrahim, A. A. AL Sahli .I. A. Alaraidh , E. M. Mostafa , and G. A. EL-Gaaly : Assessment of antioxidant activities in roots of Miswak (*Salvadora persica*) plants grown at two different locations in Saudi Arabia. Saudi J Biol Sci.; 22(2): 168–175, 2015.
- [15] K. Kamel, M. Ohtani, and M Assaf: Lignan glycosides from stems of *Salvadora persica* Phytochemistry, 31 , pp. 2469-2471, 1992.
- [16] S. S. Kim , S. Kim , E. Kim , B. Hyun, K.K .Kim and B. J. Lee: Synergistic Inhibitory Effect of Cationic Peptides and Antimicrobial Agents on the Growth of Oral Streptococci. Caries Res., 37:425–430, 2003.
- [17] I. U. Klas , P. Nicholas, P. Ankomah, F. Baquero, and R..L. Bruce . Functional relationship between bacterial density and the efficacy of antibiotics. J. Antimicrob Chemother.; 63(4): 745–757., 2009.



- [18] Wj. Loesche : Microbiology of Dental Decay and Periodontal Disease. In: Baron S, editor. Medical Microbiology. 4th edition. Galveston (TX): University of Texas Medical Branch at Galveston. Chapter 99, 1996.
- [19] M. Madigan and J. Martinko : Brock Biology of Microorganisms (11th Ed.). Prentice Hall. ISBN 0-13-144329-1 Cf. https://en.wikipedia.org/wiki/Streptococcus_mutans#cite_note-Brock-9, 2005.
- [20] A. Maguire and A. J. Rugg-Gunn. "Xylitol and Caries Prevention- British Dental Journal: 429-36, 2003.
- [21] E. S. Mahanani and S. V. Samuel: MISWAK (*Salvadora persica*) as a Cleansing Teeth. Mutiara Medika. Vol. 7 No. 1: 38-42, 2007.
- [22] P. A. Nayak , U. A. Nayak , and V. Khandelwal: The effect of xylitol on dental caries and oral flora. In Cosmet Investig Dent. 6: 89–94. 2014.
- [23] V. A. Paula, A. Modesto, K. R. Santos and R. Gleiser: Antimicrobial effects of the combination of chlorhexidine and xylitol. Br Dent J. 18; 209 (12):E19. 2010.
- [24] S. Petti S, G. Renzini , and G. Tarsitani : Effect of sucrose and glucose on the susceptibility of *Streptococcus mutans* to cephalosporins. New Microbiol.; 21(3):289-92,1998.
- [25] K. J. Ryan. and C. G. Ray (Editors) . Sherris Medical Microbiology (4th ed.). McGraw Hill. ISBN 0-8385-8529-9, 2004
- [26] A. S. Samuel, M. M. Daniel, H. Kristina, C. Heather, Y. L. Michael, B. Imran, and J. H. Richard: In Vitro Killing of Community-Associated Methicillin Resistant *Staphylococcus aureus* with Drug Combinations. Antimicrobial Agents and Chemotherapy, 48(10): 4016-4019, 2004.
- [27] S. Siddeeqh , A. Parida, M. Jose, and V. Pai : Estimation of Antimicrobial Properties of Aqueous and Alcoholic Extracts of *Salvadora Persica* (Miswak) on Oral Microbial Pathogens - An In vitro Study J. Clin Diagn Res.;10(9):p; FC13-FC16, 2016.
- [28] B. Thaweboon, S. S. Thaweboon and D. M. Tri: Fermentation of various sugars and sugar substitutes by oral microorganisms. Asian Pacific Journal of Tropical Biomedicine S258-S260, 2011.
- [29] B. K. Tiwari , V. P. Valdramidis , C. P. O'Donnell , K. Muthukumarappan , P. Bourke , and P. Cullen: Application of natural antimicrobials for food preservation. Journal of Agricultural and Food Chemistry; 57:5987, 2009.
- [30] S. M. Wåler and G. Rølla : Xylitol mechanisms of action and uses. Nor Tannlaegeforen Tid. ; 100(4):140-3, 1990.
- [31] WHO. Consensus statement on oral hygiene. Int Dent J. 50,139, 2000.
- [32] Y. Isao, T. Voichiro, K. Hirohito, O. Makiko, K. Nobuhito, H. Naoki, S. Munenori, N. Ikuo, T. Isao, M. Tatsurro, M. Hiroshi, and U. Makoto: Effect of Glucose Concentration on the pH and Growth of *Streptococcus mutans* and *Porphyromonas gingivalis*. Japanese Journal of Conservative Dentistry Vol. 58 , No. 6 p. 510-517, 2015.
- [33] Y. Zou, Y. Lee, J. Huh, and J.-W. Park: Synergistic effect of xylitol and ursolic acid combination on oral biofilms. Restor Dent Endod.; 39(4): 288–295, 2014.